

# Flow Cytometry Workshop Program

Monday 18 April 2016 - Small Operon EMBL

## Session I: (9:00-14:00) - General Knowledge

Lecture N.	Title
1	Flow Cytometry, an overview of the history and the current state of technology
2	Getting it right for Flow: Fluorochromes and Fluorescence
3	How Flow Cytometers work and generate signals: What does my cytometer tell me?
	<b>Coffee Break</b>
4	Compensation
5	Introduction into Panel Design and Sample Preparation
6	Workshop: Quizzing tutorial (Fluorochromes, matching your assay to your instrument, panel design)

Tuesday 19 April 2016 - Small Operon EMBL

## Session II: (9:00-14:00) - Specific Tools and Assays

Lecture N.	Title
1	Quality control in Flow Cytometry and adjusting cytometers to match your scientific needs
2	How does cell sorting work and what is essential to successful cell sorts
3	Cell Cycle analysis and apoptosis assays
	<b>Coffee Break</b>
4	Multicolor Flow Cytometry and the use of fluorescent proteins
5	The Basics of Imaging Flow Cytometry
6	The Basics of CyTOF
7	Workshop: Preparing for practical session

Wednesday 20 April 2016 - BD Heidelberg

## Session III: (9:00-16:00) - Practical Session at Beckman Dickinson (BD) LIMITED PLACES!

Sample Set	Experiment
1	Fluorescent proteins: CFP,mEGFP, YFP, mCherry.
2	Cell cycle measurements: PI cell cycle DAPI cell cycle 2D cell cycle (BrdU or EdU)
3	Multicolor flow: (Mouse samples, S1, fixed): Spleen

Thursday 21 April 2016 - Seminar Room EMBL

## Session IV: (9:00-14:00) - Data Analysis FLOWJO

Data from measured samples in session III will be analysed using Flowjo.

This training session is intended to give an initial approach to the software and will cover basic operations including: the creation of FlowJo workspaces, set-up of gating strategies, statistical functions, batch analyses and the generation of graphical layouts and tables.