

# Fluorescence in Flow Cytometry

Dr Malte Paulsen

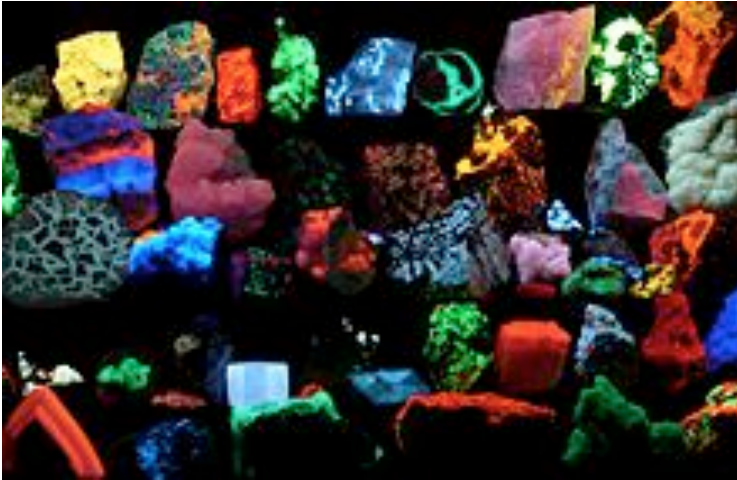
Manager, Flow Cytometry Core Facility

EMBL, Heidelberg

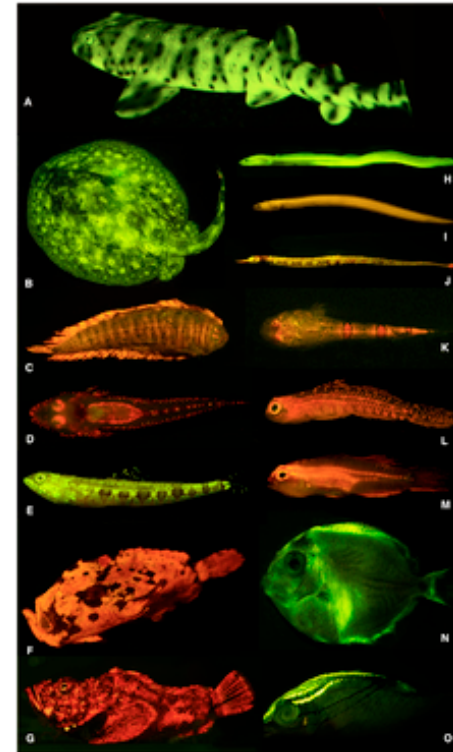
EMBL



# Fluorescence is everywhere!

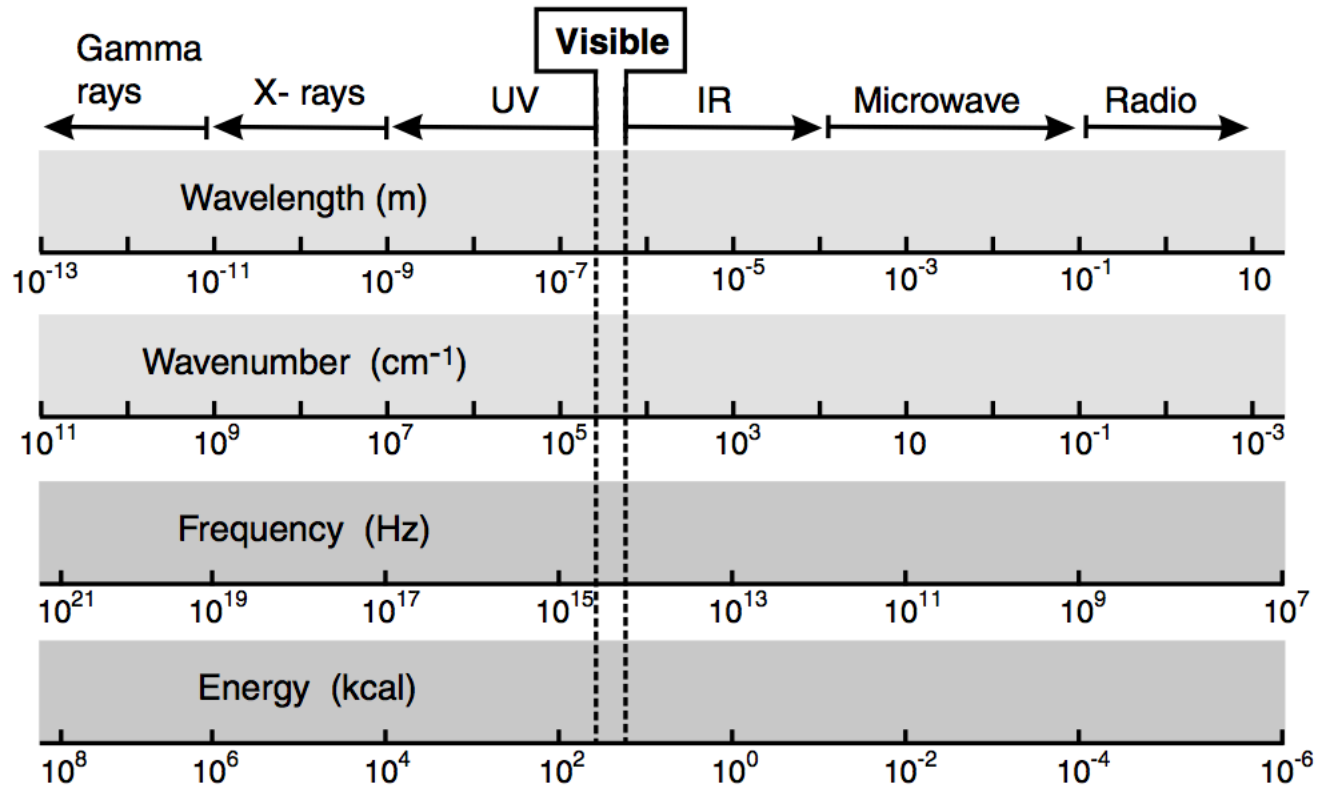


In nature...



In biotech...

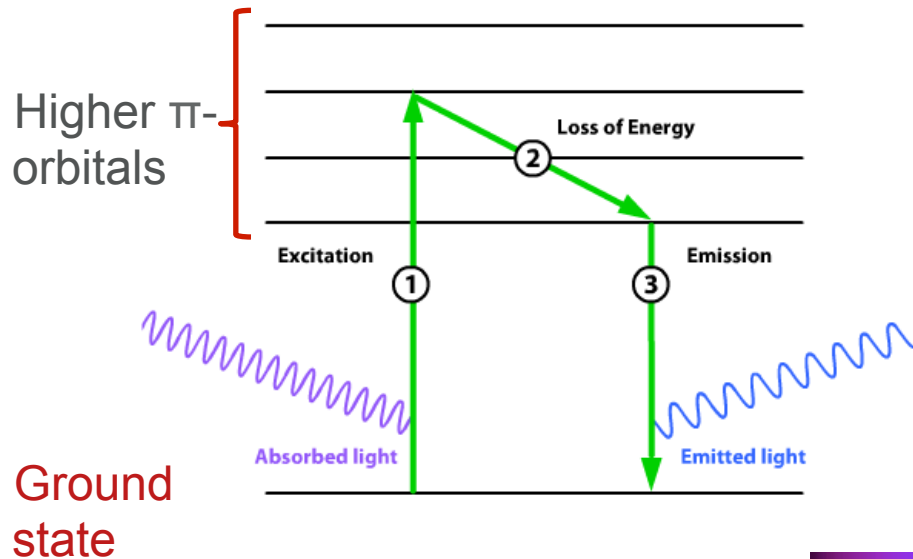
# How does Fluorescence work



Wiley, Principles of spectroscopy

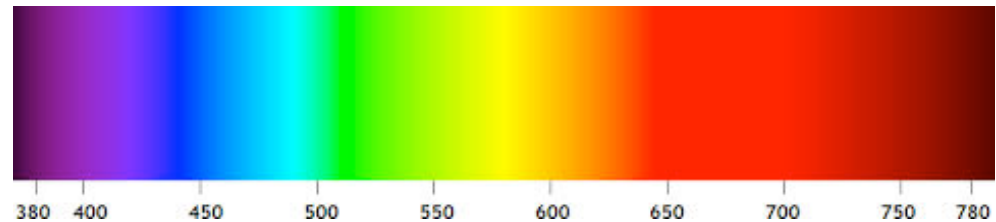
# How does Fluorescence work

## Fluorescence in a nutshell



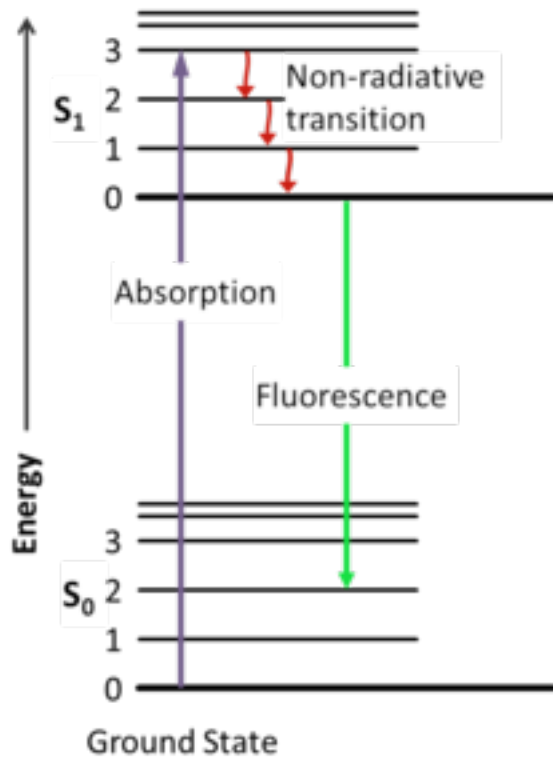
Jablonski diagram

(image source Wikipedia)

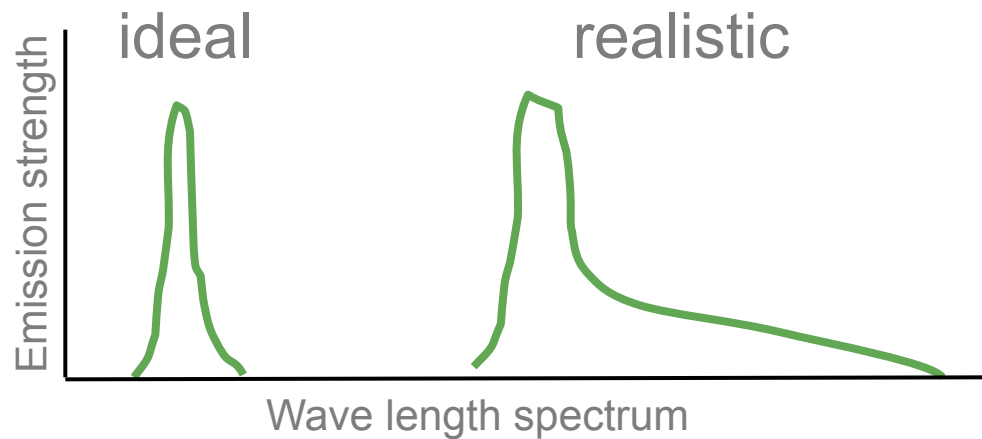




# How does Fluorescence Work



→ Depending on the amount of heat dissipation and molecule movements, the emission is never homogeneous!

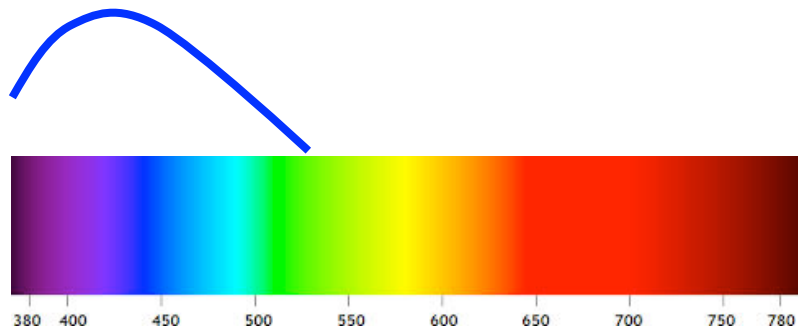


Jablonski diagram

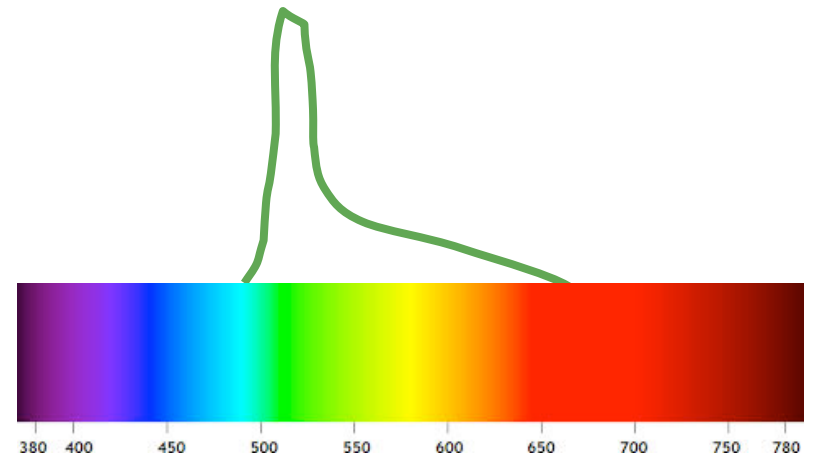
(image source Wikipedia)

# How does Fluorescence Work

- Depending on the amount of heat dissipation and molecule movements, the emission is never homogeneous!
- We see a 'distinct' fluorescence color, because this is the most abundant fluorescence emitted from the respective molecule



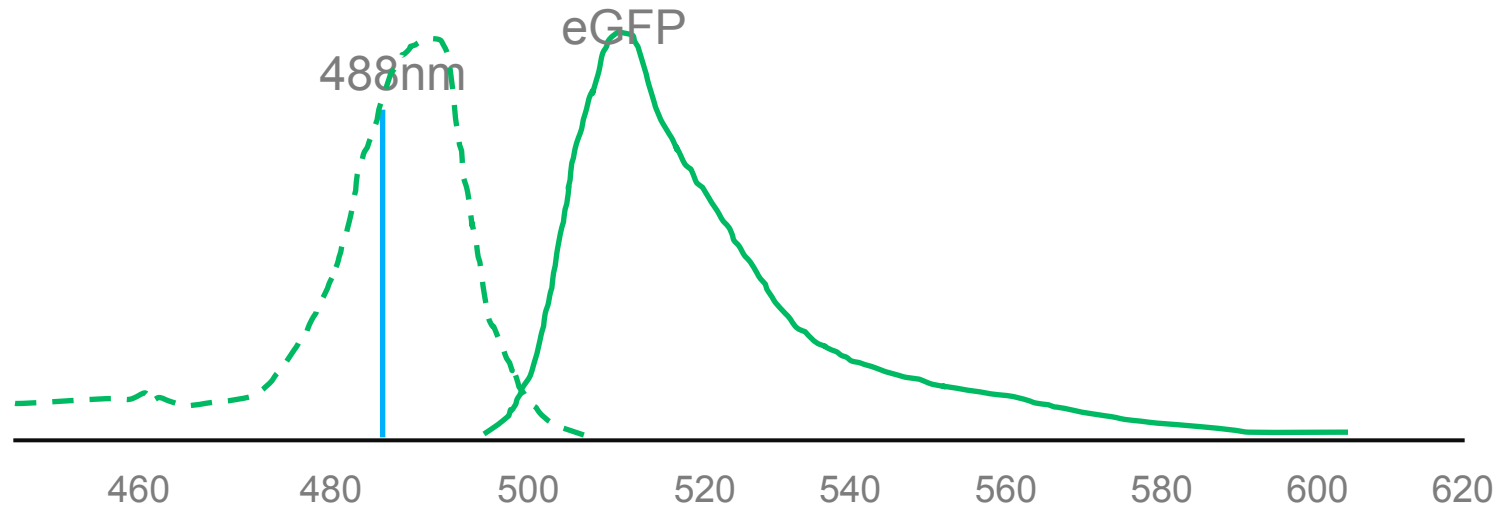
DAPI



FITC

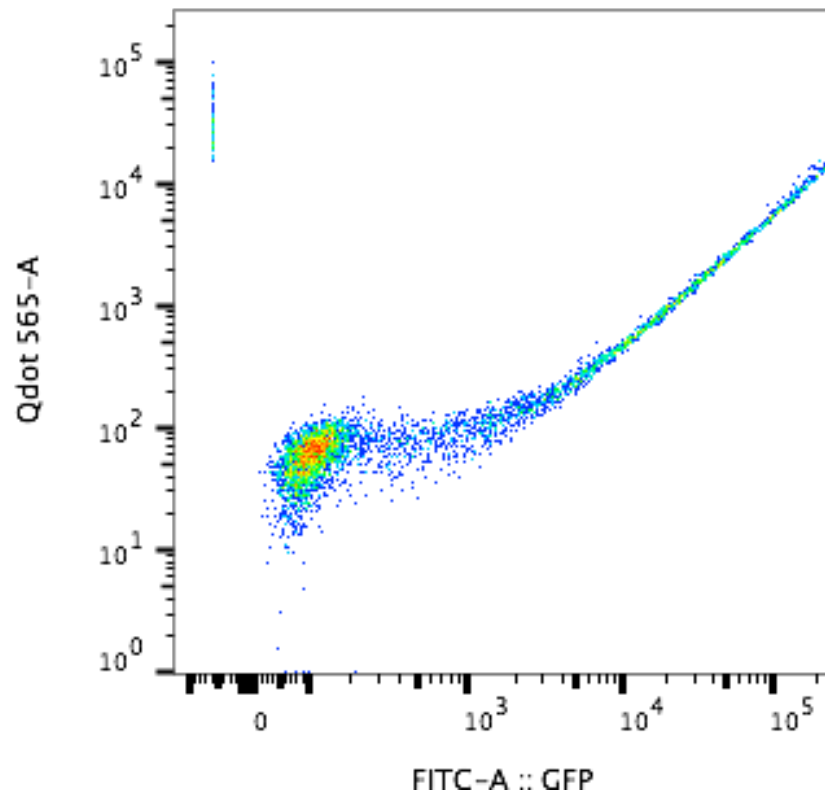
# How does Fluorescence work

→ As the emission of fluorescence is commonly biased but stochastic, how does the excitation side behave?



**Mirror-Effect: Excitation and emission behave as if they are mirrored!**

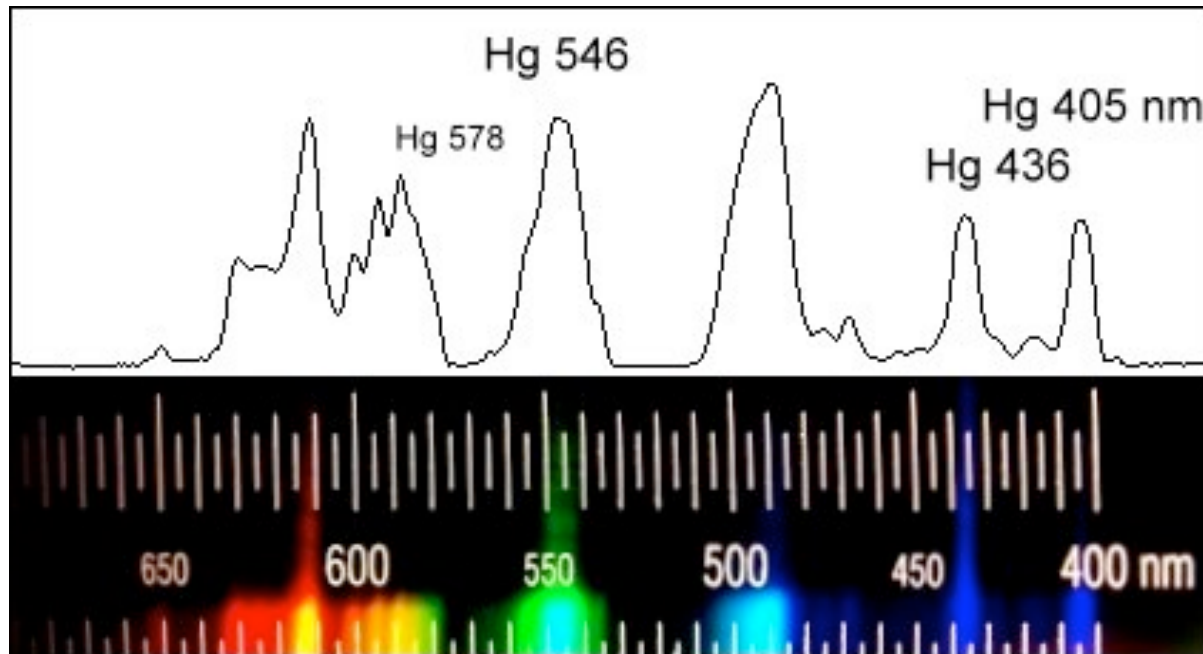
# How does Fluorescence work



Mirror-Effect: Excitation and emission  
behave as if they are mirrored!

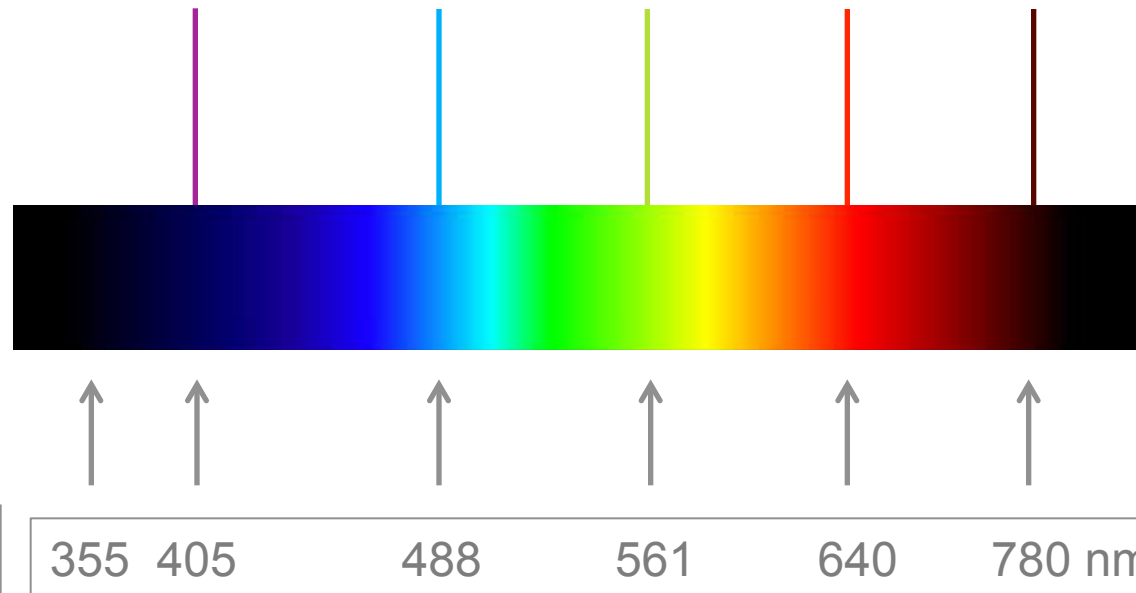
# How does Fluorescence work

Hg low pressure lamps are commonly used in simple microscopes, but they would only be mildly suitable for cytometers: too dim, too broad in their spectrum, slightly suboptimal wave lengths.



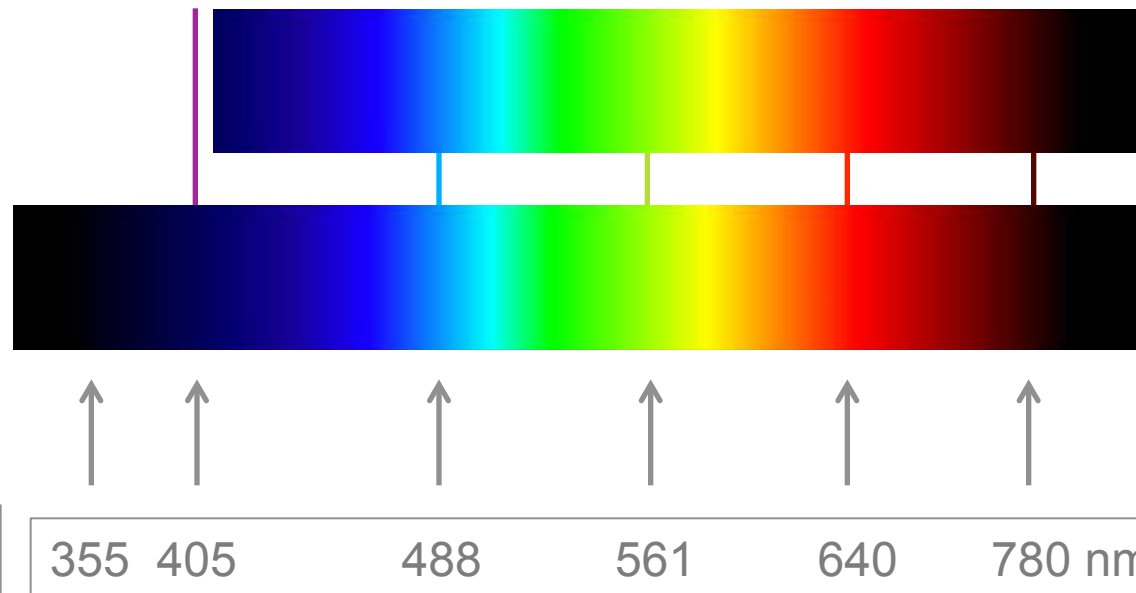
# How does Fluorescence work

To generate specificity and efficiency of excitation one uses lasers of different wave lengths



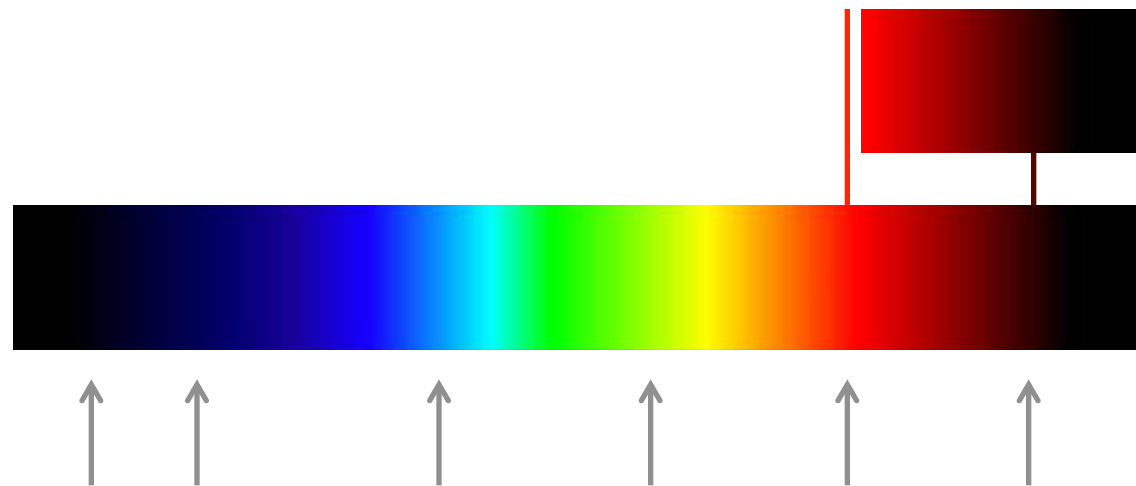
# How does Fluorescence work

As the fluorescence usually\* has a longer wavelength than the excitation, and this limits the choice of dyes that can be used.



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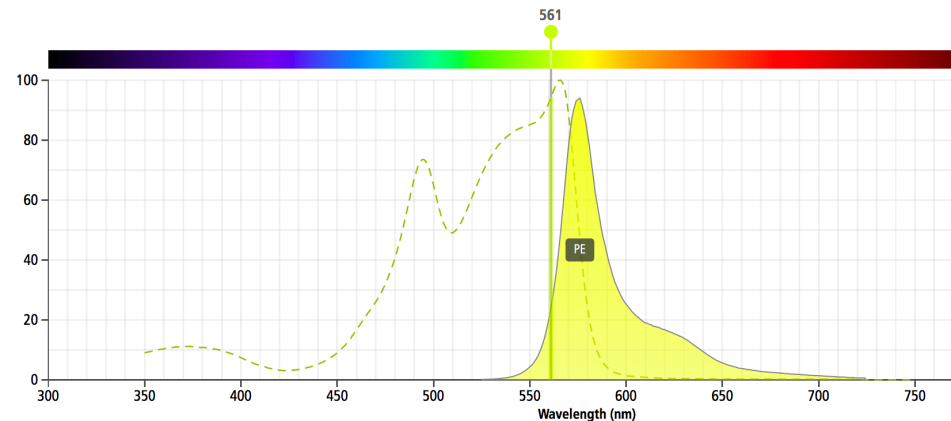
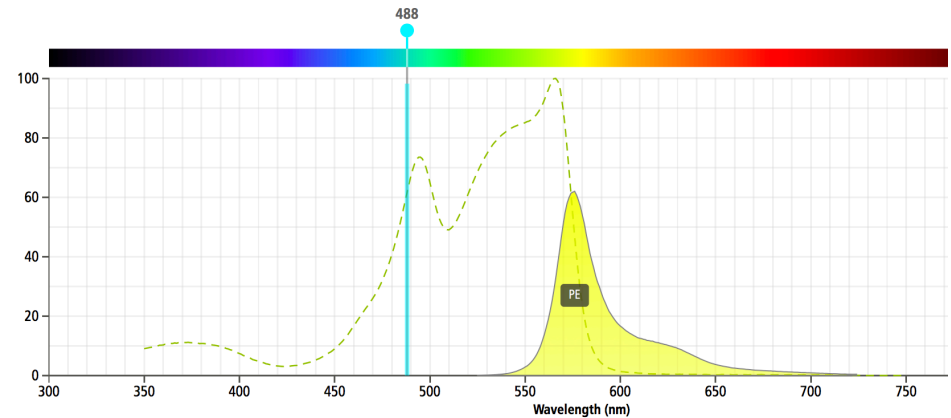
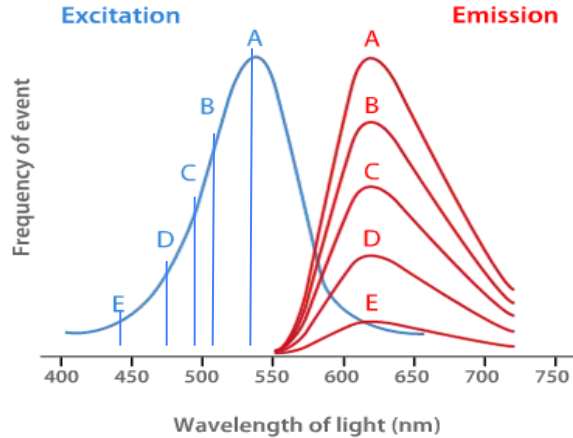
Typical laser  
wave lengths

355 405 488 561 640 780 nm



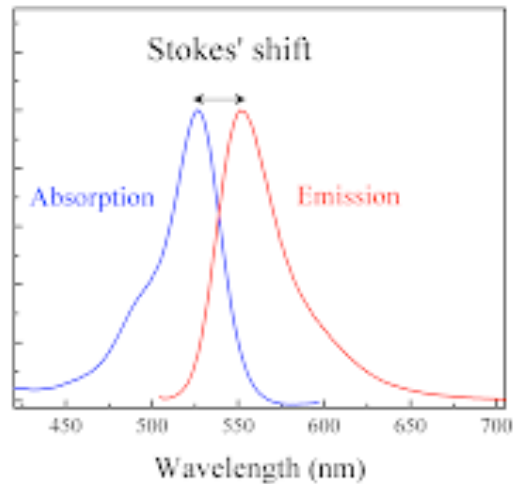
# How does Fluorescence work

The intensity of Fluorescence is coupled to the maximum excitation, but its emission spectrum will never change no matter the excitation used!



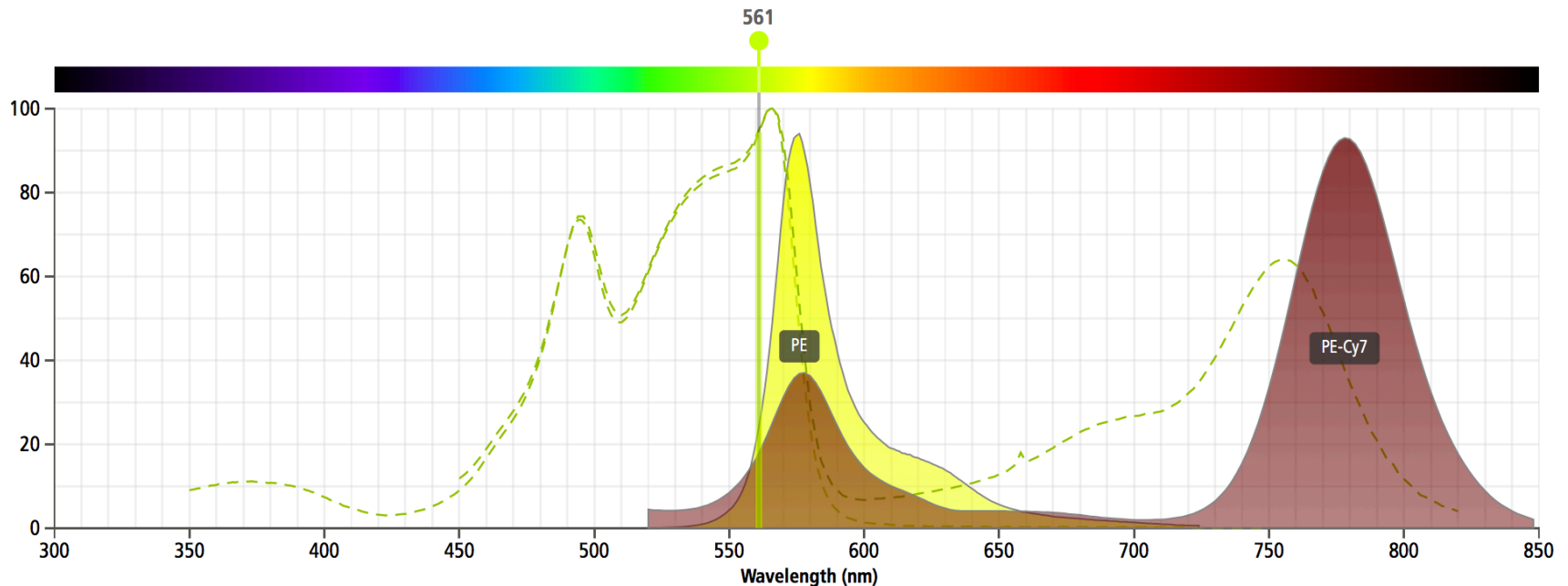
# How does Fluorescence work

The distance in nm wave length between the excitation and emission maxima is called “Stoke’s Shift”.



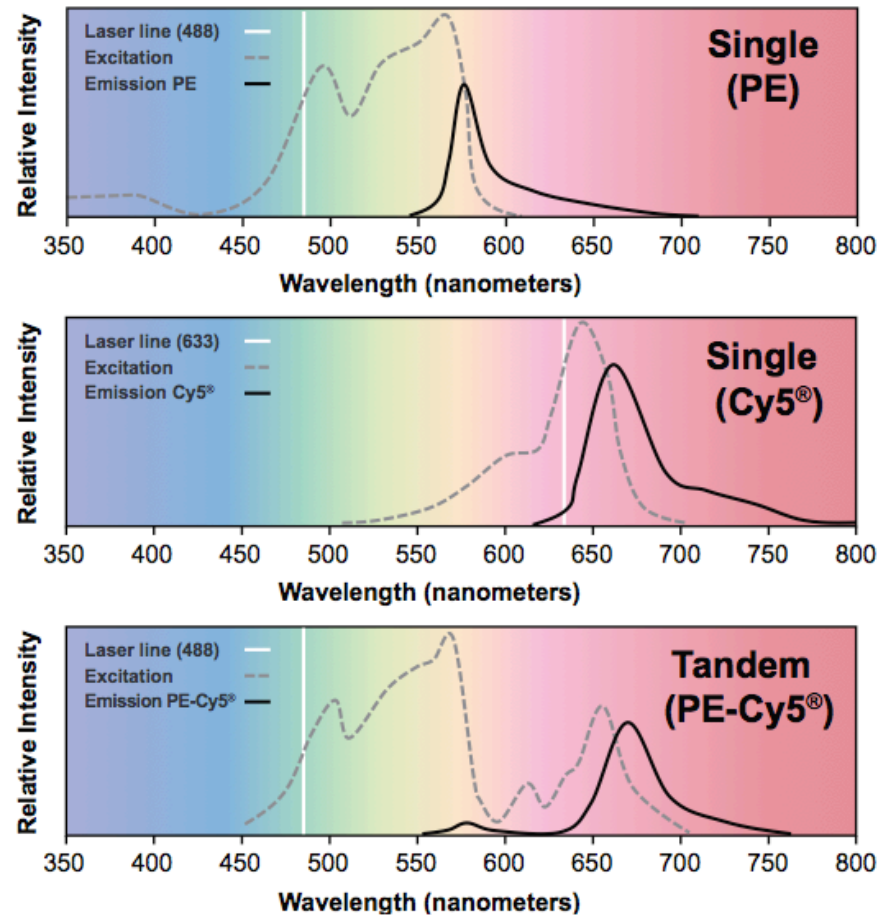
# How does Fluorescence work

The “Stoke’s Shift” can be increased by utilizing suitable FRET pairs which are known in Flow Cytometry as “Tandem Dyes”.



# How does Fluorescence work

## Excitation and Emission Spectral Profiles



Abcam website

# How does Fluorescence work

## Classical tandem dyes:

PerCP-Cy5 and PerCP-Cy5.5

PE-CF594 (PE-TexasRed)

PE-Cy5 and PE-Cy5.5

PE-Cy7

APC-R700

APC-Cy7 and APC-H7

BV605, BV655, BV711 and BV786

BUV737 and BUV800

[Abcam website](#)

# How does Fluorescence work

The brightness of a fluorochrome is based on its ability to pick up photons and to convert them into fluorescence: Quantum Efficiency/Quantum Yield



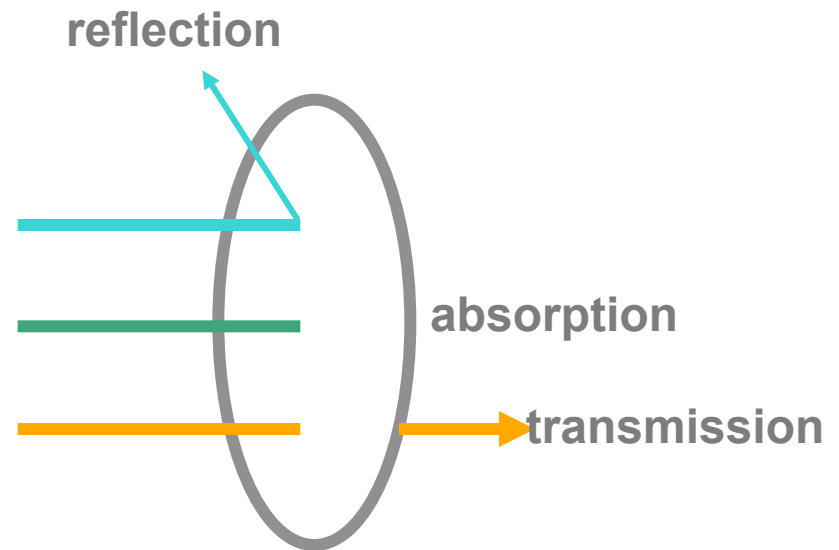
So we can formulate that dyes differ on their ability to create fluorescence: dim, mid, and bright dyes!

# How do we utilize fluorescence

...in cytometers we „pick“ the maximum of the fluorescence peaks with filters to limit the light that hits the detector and to create specificity.

**Optical filters modify the spectral distribution of the light scatter and fluorescence signals on their way to the detectors**

**An optical filter can either reflect, absorb or transmit photons**

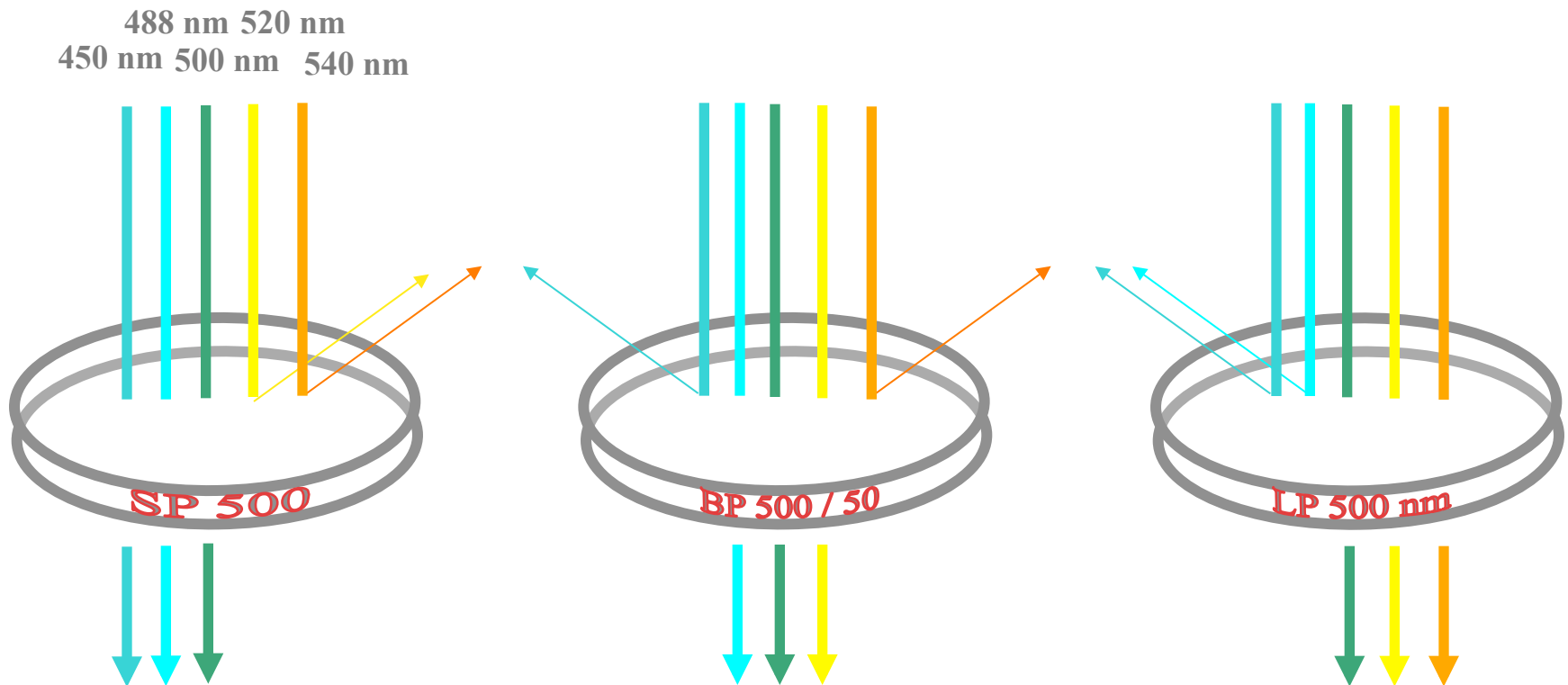


# How do we utilize fluorescence

Short Pass

Band Pass

Long Pass

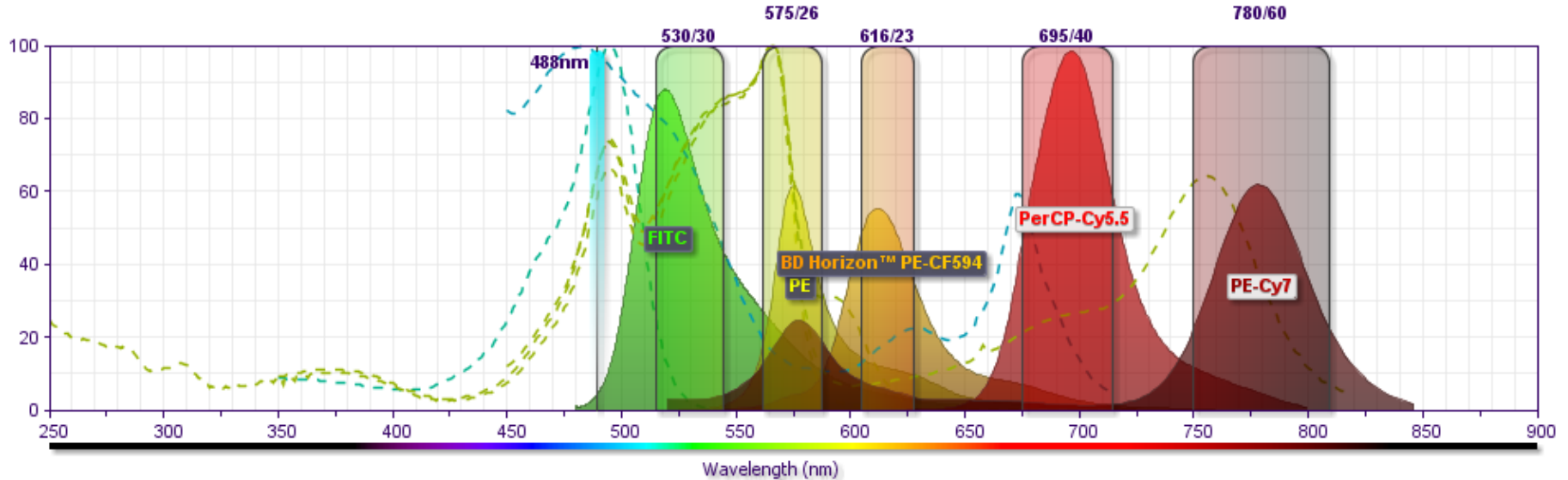


BP 500/ 50 = 475 - 525 nm



# How do we utilize fluorescence

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# How do we utilize fluorescence

## Functional dyes:

DNA binding dyes  
(DAPI Hoechst PI  
7AAD SybrGreen,  
etc)

Mitochondrial dyes  
(Mitotracker, JC-1,  
etc.)

Calcium flux dyes  
(Indo-1, etc)

...many, many  
more!

## Fluorescent proteins:

Blue (BFP, CFP)

Green (GFP, YFP,  
Venus, Citrulin)

Orange (RFP,  
mCherry, dTomato)

Red (iRFP, etc...)

Reporter cell lines  
and protein tags...

## Labeling Dyes:

“Proteins”: PE, APC,  
PerCP

Classic small  
molecules: Alexa's,  
Daylight's, FITC

Semiconductors:  
Qdots or nano  
crystals

Organic arrays:  
Brilliant (ultra) Violets

# How do we utilize fluorescence

BFP      CFP Cerulian mTurquoise  
 eGFP YFP Venus      mCherry RFP dTomato PE Plum mKate2 APC      iRFP

V450

V500  
FITC

PE

PE-CF594

PerCP  
APC

PerCP-Cy5.5  
AF700

PE-Cy7  
APC-Cy7

BV421

BV510

BV570

BV605

BV655

BV711

BV786

BUV393

BUV496

BUV563

BUV661

BUV737

BUV805

AFs →

Dylights →

Qdots →

# How do we utilize fluorescence

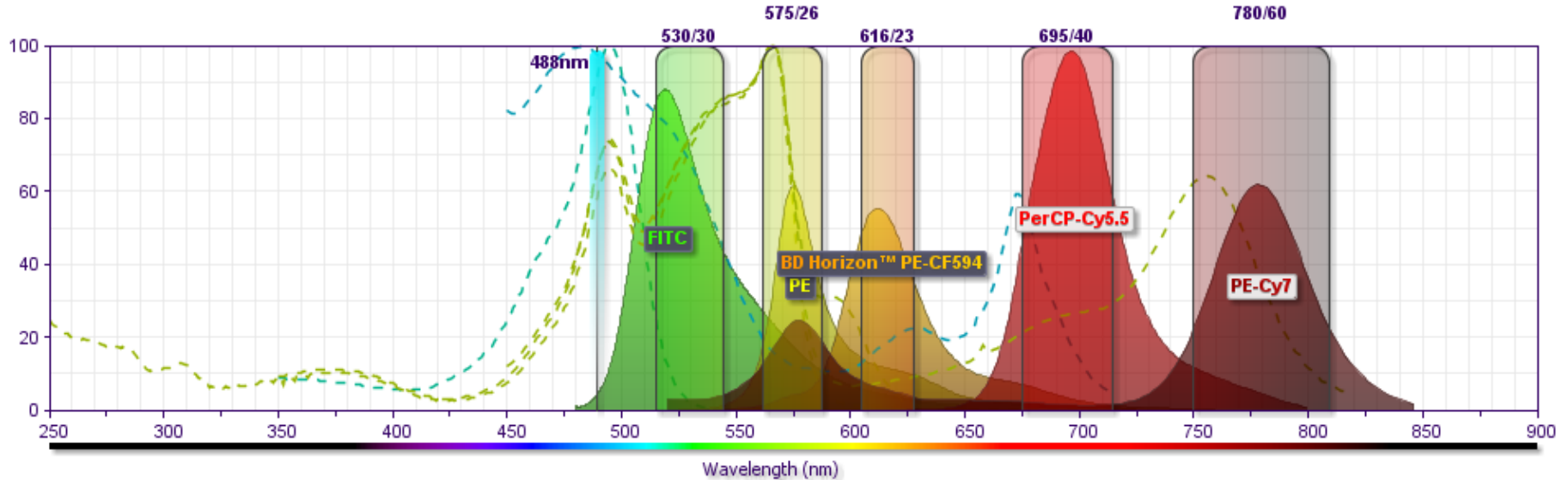
Whenever you think about setting up a FACS experiment or you want to explore options, use a spectrum viewer tool

[https://www.bdbiosciences.com/sg/research/multicolor/spectrum\\_viewer/index.jsp](https://www.bdbiosciences.com/sg/research/multicolor/spectrum_viewer/index.jsp)

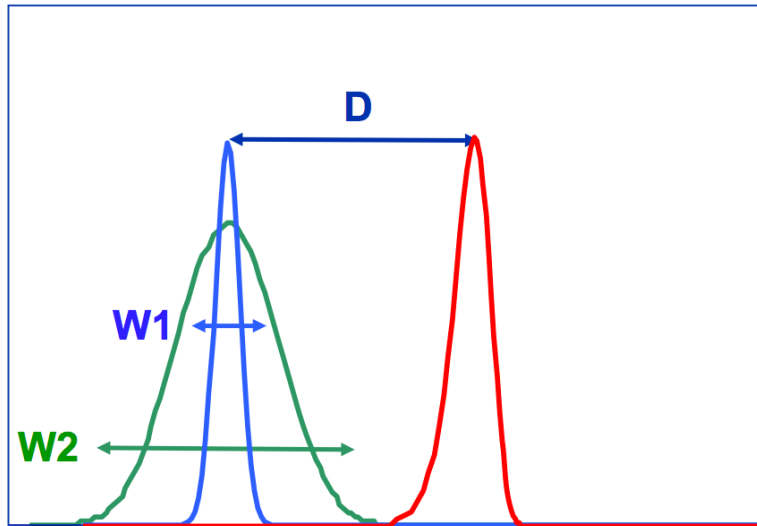
<https://www.thermofisher.com/de/de/home/life-science/cell-analysis/labeling-chemistry/fluorescence-spectraviewer.html>

# How do we utilize fluorescence

...in cytometers we „pick“ the maximum of the fluorescence peaks with filters to limit the light that hits the detector and to create specificity.



# How do we match dyes and epitope



















$$\text{Stain Index (SI)} = D/W$$

	Reagent	Clone	Filter	Stain Index
	PE	RPA-T4	575/26	305
	APC <sup>1</sup>	RPA-T4	660/20	263
	PE-Cy <sup>TM</sup> 5 <sup>2</sup>	RPA-T4	695/40	198
	Alexa Fluor® 647 <sup>1</sup>	RPA-T4	660/20	184
	PE-Cy <sup>TM</sup> 7	RPA-T4	780/60	122
	PerCP-Cy <sup>TM</sup> 5.5 <sup>2</sup>	RPA-T4	695/40	99
	Alexa Fluor® 488 <sup>3</sup>	RPA-T4	530/30	68
	BD Horizon <sup>TM</sup> V450 <sup>5</sup>	RPA-T4	450/50	65
	Alexa Fluor® 700	RPA-T4	720/40	64
	Pacific Blue <sup>TM.5</sup>	RPA-T4	450/50	63
	FITC <sup>3</sup>	RPA-T4	530/30	43
	AmCyan <sup>6</sup>	RPA-T4	525/50	37
	APC-Cy7 <sup>4</sup>	RPA-T4	780/60	36
	PerCP <sup>2</sup>	RPA-T4	695/40	30
	BD Horizon <sup>TM</sup> V500 <sup>6</sup>	RPA-T4	525/50	27
	BD APC-H7 <sup>4</sup>	RPA-T4	780/60	25

# How do we match dyes and epitope

## Chromophore brightness

	Reagent	Clone	Filter	Stain Index
	PE	RPA-T4	575/26	305
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	PE-Cy <sup>TM</sup> 5 <sup>2</sup>	RPA-T4	695/40	198
	Alexa Fluor <sup>®</sup> 647 <sup>1</sup>	RPA-T4	660/20	184
	PE-Cy <sup>TM</sup> 7	RPA-T4	780/60	122
	PerCP-Cy <sup>TM</sup> 5.5 <sup>3</sup>	RPA-T4	695/40	99
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## Antigen density

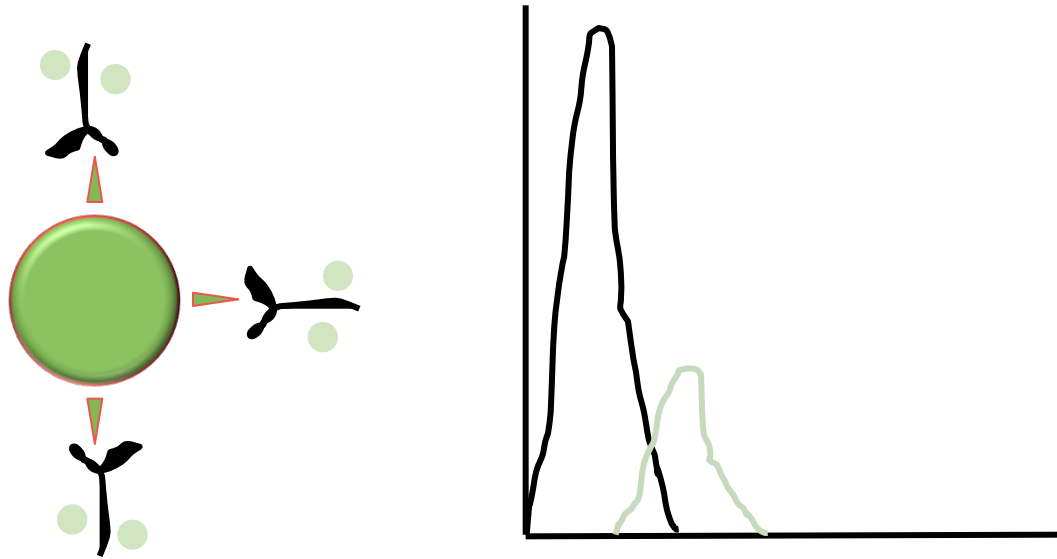
Bright dyes on lowly expressed markers!

+

Dim dyes on highly expressed markers!

# How do we utilize fluorescence

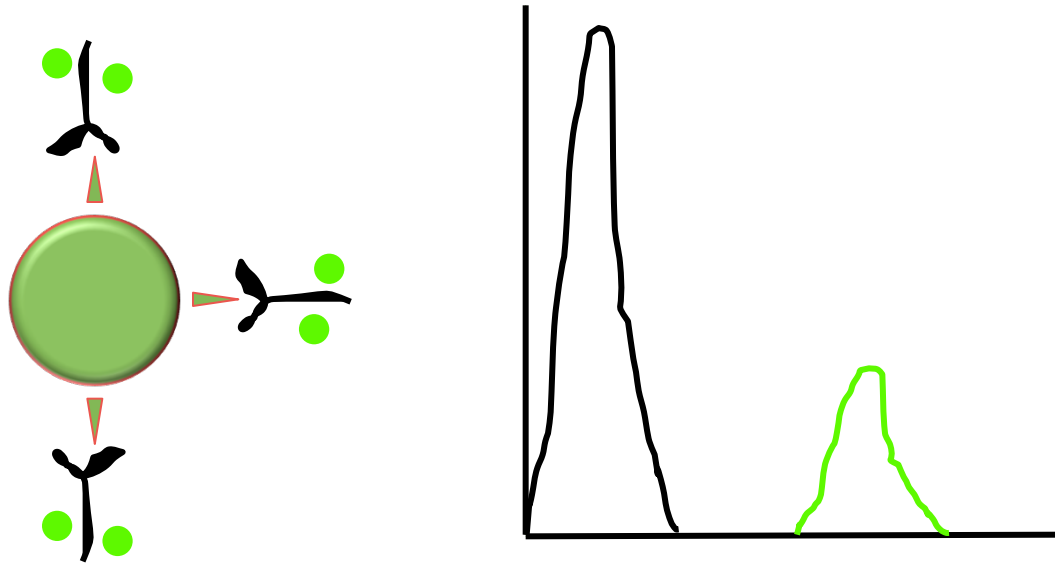
Rule of thumb: Bright dyes go on lowly expressed epitopes





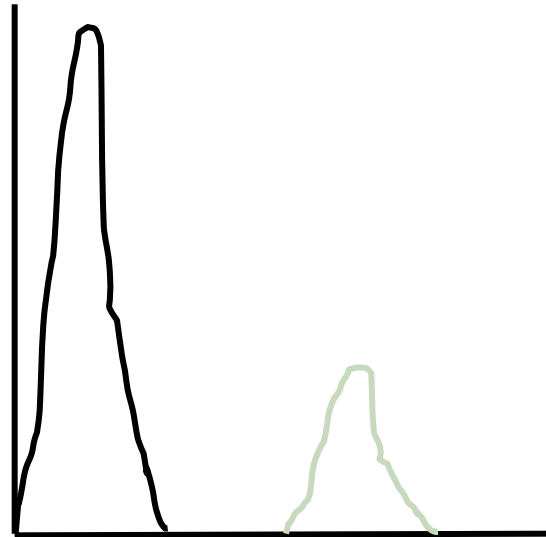
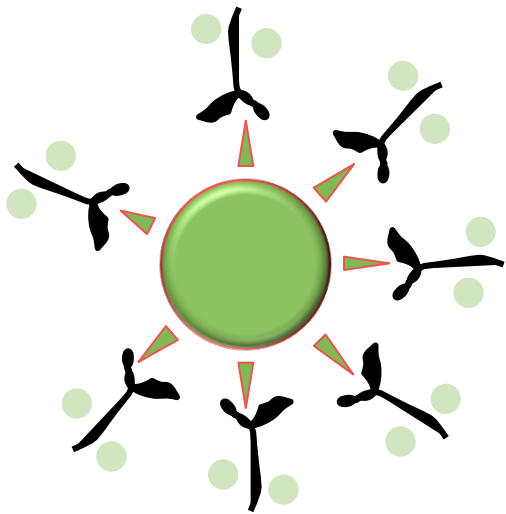
# How do we utilize fluorescence

Rule of thumb: Bright dyes go on lowly expressed epitopes



# How do we utilize fluorescence

Rule of thumb: Dim dyes go on highly expressed epitopes



# How to approach fluorescence

How many colors will I need for my experiment?

What colors are available to me (limitation on Abs)

What instrument do I have?

Will I have to use a fluorescent protein?

Can I CRISPR a HA- or Flag-Tag into my cells instead if I have a membrane protein?

Primary coupled Abs are usually better for FACS, but sometimes you need to amplify your signal.



Questions...please hit me with'em